



Kansuinone, a novel euphane-type triterpene from *Euphorbia kansui*

Jie Guo^{a,b}, Hong-Ping He^a, Xin Fang^a, Ying-Tong Di^a, Shun-Lin Li^a, Zhen Zhang^a, Ying Leng^c, Hui-Ming Hua^{b,*}, Xiao-Jiang Hao^{a,*}

^aState Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming 650204, China

^bDepartment of Natural Products Chemistry, Shenyang Pharmaceutical University, Shenyang 110016, China

^cShanghai Institute of Materia Medica, Chinese Academy of Sciences, Shanghai 201203, China

ARTICLE INFO

Article history:

Received 5 July 2010

Revised 8 September 2010

Accepted 21 September 2010

Available online 25 September 2010

Keywords:

Euphorbia kansui

Euphane-type triterpene

Structure elucidation

11 β -HSD1 inhibitory activity

ABSTRACT

Kansuinone (**1**), a rearranged euphane triterpenoid containing a spiro [5,6] ring system, was isolated from the roots of *Euphorbia kansui*. Its structure and stereochemistry were elucidated on the basis of spectroscopic, CD, and computational methods. Kansuinone (**1**) exhibited inhibitory activity against human and mouse 11 β -HSD1 (11 β -hydroxysteroid dehydrogenase type 1), with IC₅₀ values of 1.12 and 1.08 μ M, respectively.

© 2010 Elsevier Ltd. All rights reserved.

Plants of the *Euphorbia* genus produce structurally unique and diversified diterpenoids and triterpenoids, which have attracted great interest from biogenetic, synthetic, biological, and toxicological points of view.¹ Some of these compounds have shown potent antitumor, cytotoxic, antiviral, NADH oxidase inhibitory, and multidrug resistance-reversing activities.²

Euphorbia kansui is a vivacious herb distributed in the central and western parts of China, from which a novel euphane-type triterpenoid, kansuinone (**1**), and its presumed bioprecursor, kansenone (**2**) were isolated during our continuous search for biologically active compounds from the genus *Euphorbia*.

Compound **1** possessed a rearranged backbone involving a spiro [5,6] ring system that may have an origin from euphane-type triterpenoids. Previously, spirotriterpenoids have been found only as lanostane-type triterpenoids derivative.³ Compound **1** exhibited inhibitory activity against human and mouse 11 β -HSD1, with IC₅₀ values of 1.12 and 1.08 μ M, respectively. Reported here is the isolation, structure elucidation, and biological activity assays of **1**.

The roots of *E. kansui* used in this investigation were collected at Kuitun, Gansu Province, P.R. China, in December 2007. The dried and powdered roots of *E. kansui* (20 kg) were extracted with 95% EtOH. A crude residue (760 g) was obtained after evaporating the solvent and the residue was partitioned between petroleum ether and H₂O. The petroleum ether fraction (280 g) was separated by a combination of silica gel, amino silica gel, and Sephadex LH-20

columns. Final purification by silica gel column chromatography yielded kansuinone (**1**, 18 mg) and a known euphane-type triterpene, kansenone (**2**, 20 mg).⁴

Kansuinone **1**⁵ was obtained as colorless and optically active oil. Its molecular formula was determined to be C₃₀H₅₀O₃ by HR-TOF-MS (m/z [M+Na]⁺ 481.3657, calcd 481.3657) with six units of unsaturation. The IR absorption bands indicated the presence of ketone (1675 cm⁻¹) and hydroxyl (3432 cm⁻¹) groups. The hydroxyl group was also implied by the observation of a fragment ion at m/z 440 [M-H₂O]⁺ in the EI-MS. The 1D NMR spectra exhibited resonances for seven quaternary, nine methylene, six methine, and eight methyl carbons, which were assigned to a trisubstituted double bond, a ketone, two oxygenated methines, one secondary methyl, seven tertiary methyl, and two hydroxyl groups. This compound should be a tetracyclic triterpene since one olefinic group and one ketone accounted for two of the six units of unsaturation indicated by the molecular formula.

Comparison of the ¹H and ¹³C NMR data of **1** with those of kansenone (**2**), a euphane-type triterpene isolated from the same plant, revealed that both compounds were characterized by similar chemical shifts attributed to rings A and D and a monounsaturated side chain, suggesting a common structural motif they shared. The presence of the monounsaturated side chain was suggested by MS fragment ions at m/z 305 [M-side chain-D ring]⁺, 369 [M-2H₂O-69-2H]⁺, and 69 [CH₂CH=C(Me)₂]⁺.⁶ Two-dimensional NMR analysis confirmed the presence of these motifs (Fig. 1). Two hydroxyls, as required by its molecular formula and IR spectra, were located at positions C-3 and C-7 based on their chemical shifts and HMBC correlations from H-3 to Me-28 and Me-29, and

* Corresponding authors. Tel.: +86 871 522 3263; fax: +86 871 522 3070.

E-mail address: haoxj@mail.kib.ac.cn (X.-J. Hao).

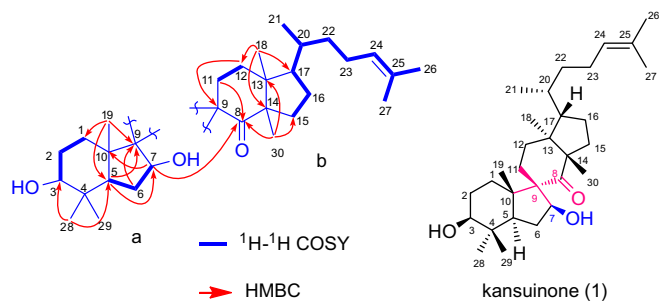


Figure 1. Selected two-dimensional NMR correlations of kansuinone (1).

H-7 to C-5. A mutually coupled system that comprised C-5, C-6, and C-7 was suggested by ^1H - ^1H COSY thus determined the structure of ring A, which fused with a contracted five-membered ring B at C-5 and C-10, forming substructure **a**, as suggested by the observation of HMBC correlations from H-5, H-6, and Me-19 to C-9 and from H-7 to C-10. Ring C, containing a ketone at C-8, was assembled by ^1H - ^1H COSY correlations between H₂-11/H₂-12 as well as HMBC correlations of H-12/C-9, H-11/C-8, Me-30/C-8, and Me-18/C-12. It fused with ring D connected by the side chain constituted the substructure **b**. The two substructures should join at C-9, forming a unique spiro [5,6] ring system, as confirmed by the HMBC cross peaks between H-7 and C-8 and the chemical shift of C-9 (δ_{C} 61.8). Therefore, a planar structure of **1** was derived, as shown in Figure 1.

The relative configuration of **1** was determined by ROESY experiments (Fig. 2) and computational methods. The large coupling constant ($J_{2,3} = 9.2$ Hz) of H-3 indicated that the hydroxyl group was oriented equatorially (β) at C-3.⁷ The relative configurations of the methyl groups and other protons in the rings A–D were ascertained on the basis of the ROESY correlations. The significant ROESY correlations of H-3/H-5, H-3/Me-28, and H-5/H-7 indicated that H-3, H-5, H-7, and Me-28 were cofacial, adopting an α -orientation. The relative configuration of C-9 was assigned by computational methods. As shown in Figure 3, the close fitting correlations between predicted and experimental chemical shifts for 9S ($R = 0.995$) than 9R-kansuinone ($R = 0.983$) (excluding that of C-8 with large chemical shift and lead to the leveling effect) indicate 9S-kansuinone is the preferred one. The 9S configuration was also consistent with its NOESY correlations, the cross-peaks of H-7/CH₃-

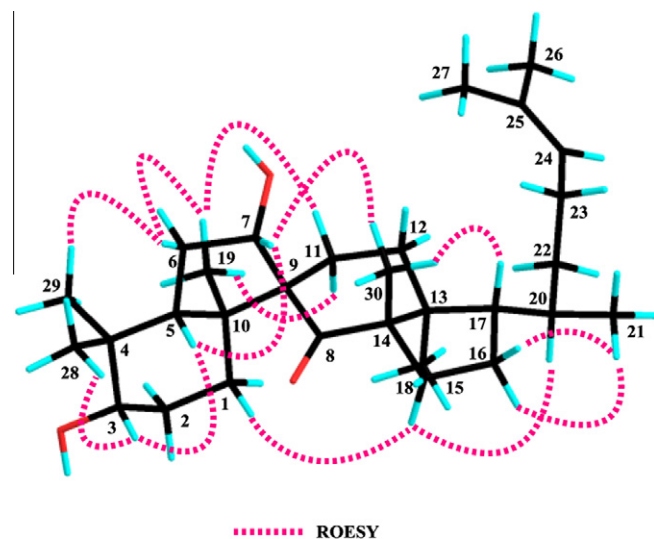


Figure 2. Key ROESY correlations of kansuinone (1).

Table 1
 ^1H , ^{13}C NMR data for kansuinone (1) in CDCl_3

	δ_{H} (multi, J in Hz)	δ_{C}
1 α	2.10 (1H, m)	29.7 (t)
1 β	1.20 (1H, m)	
2	1.67 (2H, m)	28.2 (t)
3	3.49 (1H, dd, 6.4, 9.2 Hz)	79.3 (d)
4		37.8 (s)
5	2.68 (1H, dd, 14.4, 6.0 Hz)	47.8 (d)
6 α	2.16 (1H, m)	34.7 (t)
6 β	1.43 (1H, m)	
7	4.26 (1H, t, 7.2 Hz)	76.4 (d)
8		219.3 (s)
9		61.6 (s)
10		49.0 (s)
11a	2.13 (1H, m)	24.2 (t)
11b	1.64 (1H, m)	
12a	1.88 (1H, m)	31.5 (t)
12b	1.79 (1H, m)	
13		45.9 (s)
14		61.8 (s)
15 α	1.76 (1H, m)	29.7 (t)
15 β	1.30 (1H, m)	29.7 (t)
16 α	1.31 (1H, m)	26.8 (t)
16 β	1.88 (1H, m)	
17	1.62 (1H, m)	49.6 (d)
18	0.69 (3H, s)	16.8 (q)
19	0.90 (3H, s)	17.6 (q)
20	1.46 (1H, m)	35.2 (d)
21	0.86 (3H, d, 6.4 Hz)	18.6 (q)
22 α	1.55 (1H, m)	35.3 (t)
22 β	1.13 (1H, m)	
23 α	1.88 (1H, m)	24.6 (t)
23 β	2.00 (1H, m)	
24	5.07 (1H, dd, 7.0, 1.2 Hz)	124.8 (d)
25		131.5 (s)
26	1.68 (3H, s)	25.7 (q)
27	1.60 (3H, s)	17.7 (q)
28	0.99 (3H, s)	29.7 (q)
29	0.88 (3H, s)	16.6 (q)
30	1.16 (3H, s)	22.4 (q)

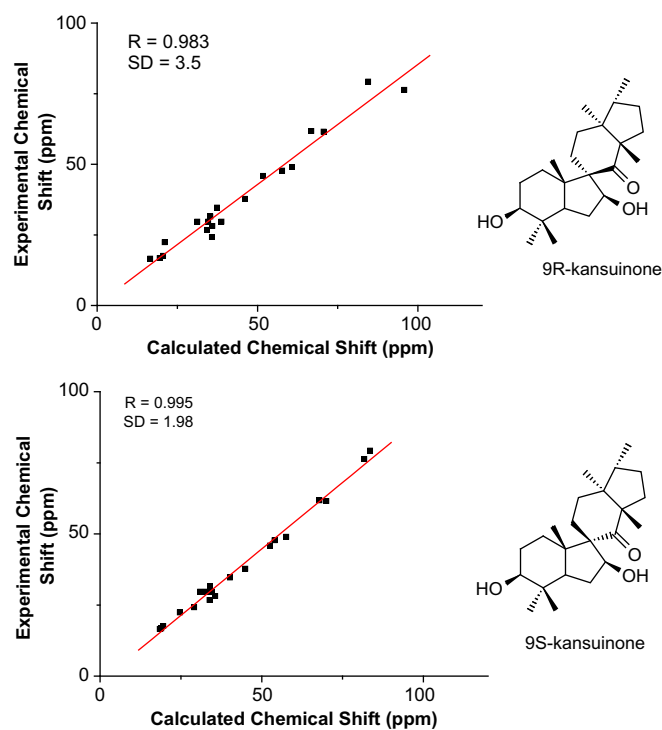


Figure 3. The correlations between the calculated and experimental chemical shifts for 9R and 9S-kansuinone by removing the point with large chemical shift (>200 ppm).

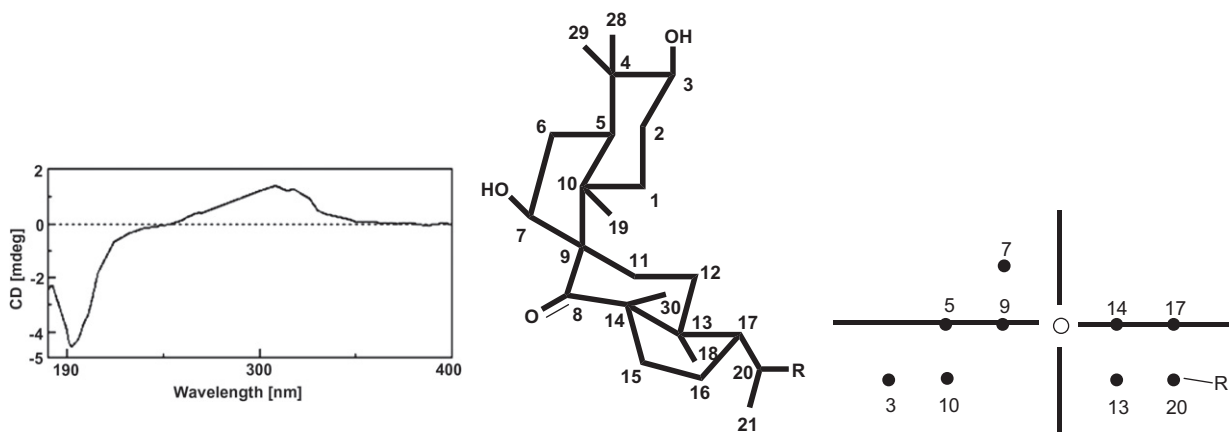


Figure 4. CD spectra and the distribution in octants of kansuione (1).

30 and CH_3 -19/ CH_2 -11. This configuration necessitated a dihedral angle near to 90° among rings A, B and C, D to maintain an anti-coplanar conformation. The ROESY cross-peaks of Me-29/ 6β , Me-19/ $\text{H}-6\beta$, CH_2 -11/Me-19, Me-30/ $\text{H}-7$, Me-18/ $\text{H}-1$, and Me-30/ $\text{H}-17$ indicated that Me-19 was β -oriented, whereas Me-30 and H-17 were oriented outside the plane defined by the C/D rings and Me-18 was directed inside the plane. Furthermore, the typical chemical shift (δ_{H} 0.86)^{6,7} of Me-21 suggested that **1** belonged to the euphane series with $20\text{H}\alpha$, which was confirmed by the correlations between Me-18/ $\text{H}-20$ and Me-21/ CH_2 -16. The stereochemistry of compound **1** was determined to be consistent with the euphane-type triterpenes.^{8,9}

The absolute configuration of **1** was assigned based on circular dichroism (CD) measurements. The CD spectrum of **1** exhibited a positive Cotton effect caused by a weak $n \rightarrow \pi^*$ transition associated with the absorption of a ketone carbonyl at 305 nm (Fig. 4). Employing the octant rule for the cyclohexanone ring, the absolute configuration of **1** was assigned as 3S, 5R, 7S, 9S, 10S, 13S, 14S, 17S, and 20R.⁹

A plausible biogenetic pathway for **1** was proposed, as shown in Scheme 1, in which a Pinacol rearrangement reaction is the key step.

Inhibition of human and mouse 11β -HSD1 enzymatic activities by the two compounds were determined by scintillation proximity assay (SPA) using microsomes containing 11β -HSD1. Compound **1**

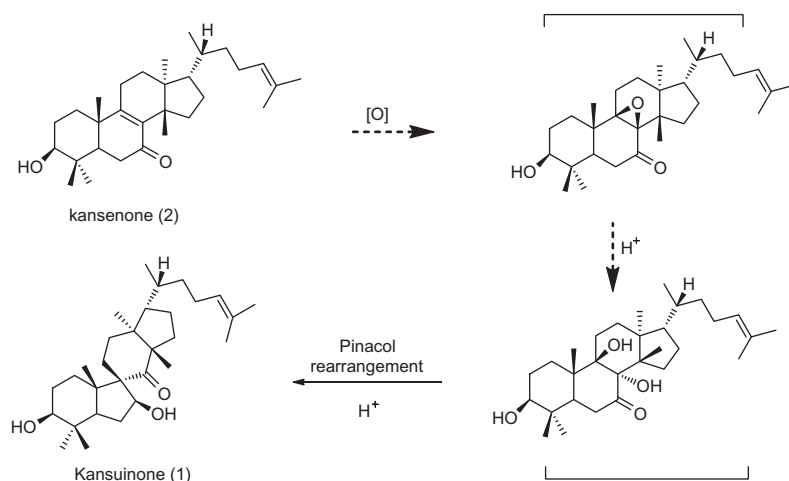
exhibited significant dose-dependent inhibition of human 11β -HSD1 with an IC_{50} of 1.12 μM and the dose-dependent inhibition of mouse 11β -HSD1 with an IC_{50} of 1.08 μM . The selectivity for the mouse HSD2/HSD1 was higher (928) than the selectivity for human HSD2/HSD1 (2.35).¹⁰

Acknowledgments

We thank Prof. Xun Gong of the Kunming Institute of Botany, Chinese Academy of Sciences, for the identification of the plant material. We also thank Prof. Jun-Min Quan, for the computations that assisted the resolution of the relative configuration. This work was financially supported by the Ministry of Science and Technology of China (2009CB940900 and 2009CB522300), National Natural Science Funding of China (21072199), Natural Science Funding of Yunnan Province (2009CD112), Foundation of Chinese Academy of Sciences to H.P. He, the Yong Academic and Technical Leader Raising Foundation of Yunnan Province to H.P. He, and Foundation of State Key Laboratory of Phytochemistry and Plant Resource in West China (P2010-ZZ01).

Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.tetlet.2010.09.099.



Scheme 1. Plausible biosynthetic pathway of **1**.

References and notes

- (a) Evans, F. J.; Taylor, S. E. *Prog. Chem. Org. Nat. Prod.* **1983**, *44*, 1–99; (b) Günther, G.; Martinek, T.; Dombi, G.; Hohmann, J.; Vasas, A. *Mag. Res. Chem.* **1999**, *37*, 365–370; (c) Appendino, G.; Belloro, E.; Tron, G. C.; Jakupovic, J.; Ballero, M. *J. Nat. Prod.* **1999**, *62*, 1399–1404; (d) Vogt, G.; Mattes, E.; Rothenburger, J.; Hertkorn, N.; Achatz, S.; Sanderman, H. *Phytochemistry* **1999**, *51*, 289–295; (e) He, W. D.; Cik, M.; Lesage, A.; Linden, I. V. D.; Kinpe, N. D.; Appendino, G.; Bracke, J.; Mathenge, S. G.; Mudida, F. P.; Leysen, J. E.; Puyvelde, L. V. *J. Nat. Prod.* **2000**, *63*, 1185–1187.
- (a) Sahai, R.; Rastogi, R. P.; Jakupovic, J.; Bohlmann, F. *Phytochemistry* **1981**, *20*, 1665–1667; (b) Miglietta, A.; Gabriel, L.; Appendino, G.; Bocca, C. *Cancer Chemother. Pharmacol.* **2003**, *51*, 67–74; (c) Mucsi, I.; Molnar, J.; Hohmann, J.; Redei, D. *Planta Med.* **2001**, *67*, 672–674; (d) Basfjzl, G. L.; Checa, J.; Marco, J. A.; Estornell, E. *Planta Med.* **2003**, *69*, 177–178; (e) Hohmann, J.; Molnar, J.; Redei, D.; Evanics, F.; Forgo, P.; Kaman, A.; Argay, G.; Szabo, P. *J. Med. Chem.* **2002**, *45*, 2425–2431.
- (a) Tanaka, R.; Wada, S. I.; Aoki, H.; Matsunaga, S.; Yamori, T. *Helv. Chim. Acta* **2004**, *87*, 240–249; (b) Daoubi, M.; Marquez, N.; Mazoir, N.; Benharref, A.; Hernandez-Galan, R.; Munoz, E.; Collado, I. G. *Bioorg. Med. Chem.* **2007**, *15*, 4577–4584; (c) Vieira, L. M. M.; Kijjoo, A.; Wilairat, R.; Nascimento, M. S. J.; Gales, L.; Damas, A. M.; Silva, A. M. S.; Mondranondra, I. O.; Herz, W. *J. Nat. Prod.* **2004**, *67*, 2043–2047.
- Wang, L. Y.; Wang, N. L.; Yao, X. S.; Miyata, S.; Kitanaka, Susumu *J. Nat. Prod.* **2003**, *66*, 630–633.
- Kansuione*: colorless oiliness; $[\alpha]_D^{16} +12.4^\circ$ (c 0.19, MeOH); UV (MeOH) λ_{\max} nm 202.2; IR (KBr) ν_{\max} : 3432, 2963, 2928, 1675, 1629, 1460, 1378, 1022 and 584 cm^{-1} ; ^1H and ^{13}C NMR data (Table 1); EIMS m/z 458 $[\text{M}]^+$, 440 $[\text{M}-\text{H}_2\text{O}]^+$, 369, 305, 69; HRTOFMS m/z 481.3657 (calcd for $[\text{M}+\text{Na}]^+$ 481.3657).
- (a) Arai, Y.; Hirohara, M.; Ageta, H. *Tetrahedron Lett.* **1989**, *30*, 7209–7212; (b) Abe, I.; Rohmer, M. *J. Chem. Soc., Perkin Trans. 1* **1994**, *7*, 783–791; (c) Mamta, M.; Yogendra, N.; Sushil, K. *Phytochemistry* **2000**, *54*, 835–838.
- (a) Jiang, Z. H.; Tanaka, T.; Hirata, H.; Fukuoka, R.; Kouno, I. *Tetrahedron* **1997**, *16999–17008*; (b) Akihisa, T.; Kimura, Y.; Kokke, W. C. M. C.; Takase, S.; Yasukawa, K.; Tamura, T. *J. Chem. Soc., Perkin Trans. 1* **1996**, 2379–2384.
- David Nes, W.; Wong, R. Y.; Benson, M.; Landery, J. R.; Nes, W. R. *Proc. Natl. Acad. Sci.* **1984**, *81*, 5896–5900.
- Moffitt, W.; Woodward, R. B.; Moscovitz, A.; Klyne, W.; Djerassi, C. *J. Am. Chem. Soc.* **1961**, *83*, 4013–4018.
- (a) Hanson, R. W.; Reshef, L. *Annu. Rev. Biochem.* **1997**, *66*, 581–611; (b) Bujalska, I. J.; Kumar, S.; Hewison, M.; Stewart, P. M. *Endocrinology* **1999**, *140*, 3188–3196; (c) Masuzaki, H.; Paterson, J. M.; Shinyama, H.; Morton, N. M.; Mullins, J. J.; Seckl, J. R., et al *Science* **2001**, *294*, 2166–2170; (d) Paterson, J. M.; Morton, N. M.; Fievet, C.; Kenyon, C. J.; Holmes, M. C.; Staels, B., et al *Proc. Natl. Acad. Sci.* **2004**, *101*, 7088–7093; (e) Yang, H. Y.; Shen, Y.; Chen, J. H.; Jiang, Q. F.; Leng, Y.; Shen, J. H. *Eur. J. Med. Chem.* **2009**, *44*, 1167–1171; (f) Kotelevtsev, Y.; Holmes, M. C.; Burchell, A.; Houston, P. M.; Schmoll, D.; Jamieson, P. *Proc. Natl. Acad. Sci.* **1997**, *94*, 14924–14929.